



# EvenCoat™ Goat Anti-mouse IgG Microplates

## ORDERING INFORMATION & SPECIFICATIONS

**Catalog Numbers:** CP001  
CP002

**Sizes:** 5 Coated Microplates (Catalog # CP001)  
15 Coated Microplates (Catalog # CP002)

**Storage:** 2 - 8° C\*

**Coating Antibody:** Goat anti-mouse IgG

**Specificity:** Mouse IgG Fc Region

**Coating Volume:** 100 µL/well

**Blocking Volume:** 300 µL/well

**Suggested mouse IgG Conc.:** 1.0 - 5.0 µg/mL  
(0.1 - 0.5 µg/well)

## Intended Use

EvenCoat™ goat anti-mouse IgG microplates are clear, pre-blocked 96-well polystyrene microplates coated with goat-derived antibody specific for the Fc region of mouse IgG. These plates may be used as a solid support for most sandwich ELISAs utilizing a mouse IgG capture antibody and a non-mouse IgG detection antibody. Other applications include competitive ELISA, IgG isotyping, and hybridoma screening/selection. An example protocol for a sandwich ELISA is described below.

## Related Products

- Stop Solution (Catalog # DY994)
- Streptavidin-HRP (Catalog # DY998)
- Substrate Solution (Catalog # DY999)
- Wash Buffer (Catalog # WA126)

## Suggested Protocol

**Note:** *EvenCoat Microplates may be used in most sandwich ELISAs that use a mouse IgG capture antibody. Each capture antibody should be validated separately for use in this application, and the detection antibody should not be a mouse IgG. This protocol is for sandwich ELISAs that use a biotinylated detection antibody. Other formats may also be used.*

Reconstitute the ELISA reagents, prepare the working concentrations, and prepare the ancillary reagents per the manufacturer's specifications. EvenCoat plates are pre-blocked, so a blocking step is not necessary.

1. Add 100 µL of diluted Capture Antibody to each well. Cover with an adhesive strip, and incubate for 1 hour at room temperature.
2. Aspirate each well, and wash with Wash Buffer, repeating the process two times for a total of three washes. Wash by filling each well with Wash Buffer (400 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate, and blot it against clean paper towels.
3. Add 100 µL of sample or standard to each well. Cover with an adhesive strip, and incubate for 1.5 - 2.0 hours at room temperature.
4. Repeat the aspiration/wash as in step 2.
5. Add 100 µL of diluted Detection Antibody to each well. Cover with an adhesive strip, and incubate for 1.5 - 2.0 hours at room temperature.
6. Repeat the aspiration/wash as in step 2.
7. Add 100 µL of the working dilution of Streptavidin-HRP to each well. Cover with an adhesive strip, and incubate for 20 minutes at room temperature.

**Note:** *Streptavidin-HRP concentration may have to be adjusted for optimal signal generation.*

8. Repeat the aspiration/wash as in step 2.
9. Add 100 µL of TMB Substrate Solution to each well. Incubate for 20 minutes at room temperature. **Protect from light.**
10. Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
11. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

\*Unopened plates may be stored at 2 - 8° C. Upon opening, return unused wells to the foil pouch containing the desiccant pack, and reseal along the entire edge of zip-seal. Plates may then be stored for up to 1 month at 2 - 8° C.

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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