



## Monoclonal Anti-human XCL1/Lymphotactin Antibody

### ORDERING INFORMATION

**Catalog Number:** MAB6951

**Clone:** 109001

**Lot Number:** DVV01

**Size:** 500 µg

**Formulation:** 0.2 µm filtered solution in PBS  
with 5% trehalose

**Storage:** -20° C

**Reconstitution:** sterile PBS

**Specificity:** rhXCL1

**Immunogen:** *E. coli*-derived rhXCL1

**Ig class:** mouse IgG<sub>2B</sub>

**Applications:** Western blot  
ELISA capture

### Preparation

This antibody was produced from a murine hybridoma elicited from a mouse immunized with purified, *E. coli*-derived, recombinant human XCL1 (rhXCL1). The IgG fraction of the ascites fluid was purified by Protein G affinity chromatography.

### Formulation

Lyophilized from a 0.2 µm filtered solution in phosphate-buffered saline (PBS) with 5% trehalose.

### Endotoxin Level

< 0.1 EU per 1 µg of the antibody as determined by the LAL method.

### Reconstitution

Reconstitute with sterile PBS. If 1 mL of PBS is used, the antibody concentration will be 500 µg/mL.

### Storage

Lyophilized samples are stable for twelve months from date of receipt when stored at -20° C to -70° C. Upon reconstitution, the antibody can be stored at 2° - 8° C for 1 month without detectable loss of activity. Reconstituted antibody can also be aliquotted and stored frozen at -20° C to -70° C **in a manual defrost freezer** for six months without detectable loss of activity. **Avoid repeated freeze-thaw cycles.**

### Specificity

This antibody has been selected for its ability to recognize rhXCL1 in ELISAs and Western blots.

### Applications

**Western Blot** - This antibody can be used at 1 - 2 µg/mL with the appropriate secondary reagents to detect human XCL1. The detection limit for rhXCL1 is approximately 2 ng/lane under non-reducing and reducing conditions.

**ELISA Capture** - This antibody can be used as a capture antibody in a human XCL1 ELISA in combination with biotinylated, human XCL1 affinity purified polyclonal detection antibody (Catalog # BAF695). A general protocol is provided on the next page. Using plates coated with 100 µL/well of the capture antibody at 2 µg/mL, in combination with 100 µL/well of the detection antibody at 200 ng/mL, an ELISA for sample volumes of 100 µL with a range of 62.5 - 4000 pg/mL can be obtained. In this format, less than 0.15% cross-reactivity with rmMCP-5, rhMIP-3β, rhGROα, rhTGFα, rhGROγ, rhActivin RIIA/Fc Chimera, rhIGF-II and rhIL-15 was observed. Titrate each preparation of the recombinant protein for standard preparation to arrive at the most suitable dose range. For this antibody pair, a two-fold dilution series starting at 8 ng/mL is suggested.

**Optimal dilutions should be determined by each laboratory for each application.**

## *ELISA Protocol*

### **Solutions Required**

- **Wash Buffer** - 0.05% Tween 20 in PBS, pH 7.4
- **Diluent** - 0.1% BSA, 0.05% Tween 20 in Tris-buffered Saline pH 7.3 (20 mM Trizma base, 150 mM NaCl)
- **Substrate Solution** - 1:1 mixture of Color Reagent A (H<sub>2</sub>O<sub>2</sub>) and Color Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY998)
- **Stop Solution** - 1 M H<sub>2</sub>SO<sub>4</sub>

### **Plate Preparation**

1. Transfer 100 µL/well of the capture antibody (diluted to the appropriate concentration in PBS, use immediately) to an ELISA plate. Seal plate and incubate overnight at room temperature.
2. Aspirate each well and wash with Wash buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 µL) using a squirt bottle, multi-channel pipette, manifold dispenser or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper toweling.
3. Block plates by adding 300 µL of PBS containing 1% BSA, 5% sucrose and 0.05% NaN<sub>3</sub> to each well. Incubate at room temperature for a minimum of 1 hour.
4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition. Alternatively, the blocking buffer can be aspirated after step 3 and the plates can be dried under vacuum. When sealed with desiccant, the plates can be stored at 4° - 8° C for at least 2 months.

### **Assay Procedure**

1. Dilutions of unknowns and standards should be carried out in polypropylene tubes. Add 100 µL of sample or standards in an appropriate diluent, per well. Mix by gently tapping the plate frame for 1 minute. Cover with an adhesive strip and incubate 2 hours at room temperature.
2. Repeat the aspiration/wash as in step 2 of Plate Preparation.
3. Add 100 µL of the biotinylated detection antibody, diluted in the appropriate diluent, to each well. Cover with a new adhesive strip and incubate 2 hours at room temperature.
4. Repeat the aspiration/wash as in step 2 of Plate Preparation.
5. Add 100 µL streptavidin HRP (R&D Systems, Catalog # DY999, dilute according to the directions on the vial label) to each well. Cover the plate and incubate for 20 minutes at room temperature.
6. Repeat the aspiration/wash as in step 2.
7. Add 100 µL of Substrate Solution to each well. Incubate for 20 - 30 minutes at room temperature. Avoid placing the plate in direct light.
8. Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well within 30 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

### **Calculation of Results**

To calculate assay results, average the duplicate readings and subtract the zero standard optical density from the sample optical density. Create a standard curve using data reduction software capable of generating a four parameter (4P-L) curve fit. Alternatively, plot the optical density for the standards versus the concentration of the standards and draw the best curve. The data can be linearized by using log-log paper and regression analysis may be applied to the log transformation. To determine the human XCL1 concentrations for each sample, first find the absorbance value on the y-axis and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the x-axis and read the corresponding human XCL1 concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor. A standard curve should be generated for each set of samples assayed.

### **Limitations**

It is important that the diluents selected for reconstitution and for dilution of the standard reflect the environment of the samples being measured. The diluent suggested in the above protocol may be suitable for most cell culture supernate samples. Validate diluents for specific sample types prior to use.

A basic understanding of immunoassay development is required for the successful use of these reagents in immunoassays. The protocol provided is for demonstration purposes only. The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunoassay with a different sensitivity and dynamic range.